8. REFINEMENT OF STRUCTURAL PARAMETERS

the case of shape constraints, the equations are nonlinear, and the elements of C must be re-evaluated in each iteration.

8.3.2. Stereochemically restrained least-squares refinement

The precision with which an approximately correct model can be refined to describe the atomic structure of a crystal depends on the ability of the model to represent the atomic distributions and on the quality of the observational data being fitted with the model. In addition, although the structure can in principle be determined by a well chosen data set only a little larger than the number of parameters to be determined (Section 8.4.4), in practice, with a nonlinear model as complex as that for a macromolecular crystal, it is necessary for the parameters defining the model to be very much overdetermined by the observations. For well ordered crystals of small- and intermediate-sized molecules, it is usually possible to measure a hundred or more independent Bragg reflections for each atom in the asymmetric unit. When the model contains three position parameters and six atomic displacement parameters for each atom, the over-determinacy ratio is still greater than ten to one. In such instances, each model parameter can usually be quite well determined, and will provide an accurate representation of the average structure in the crystal, except in regions where ellipsoids are not adequate descriptions of the atomic distributions. This contrasts sharply with studies of biological macromolecules, in which positional disorder and thermal motion in large regions, if not the entire molecule, often limit the number of independent reflections in the data set to fewer than the number of parameters necessary to define the distributions of individual atoms. This problem may be overcome either by reducing the number of parameters describing the model or by increasing the number of independent observations. Both approaches utilize knowledge of stereochemistry.

A great deal of geometrical information with which an accurate model must be consistent is available at the onset of a refinement. The connectivity of the atoms is generally known, either from the approximately correct Fourier maps of the electron density obtained from a trial structure determination or from sequencing studies of the molecules. Quite tight bounds are placed on local geometry by the accumulating body of information concerning bond lengths, bond angles, group planarity, and conformational preferences in torsion angles. Additional knowledge concerns van der Waals contact potential functions and hydrogen-bonding properties, and displacement factors must also be correlated in a manner consistent with the known geometry. In Section 8.3.1, we discuss the use of constraints to introduce this stereochemical knowledge. In this section, we discuss a technique that introduces the stereochemical conditions as additional observational equations (Waser, 1963). This method differs from the other in that information is introduced in the form of distributions about mean values rather than as rigidly fixed geometries. The parameters are restrained to fall within energetically permissible bounds.

8.3.2.1. Stereochemical constraints as observational equations

As described in Section 8.1.2, given a set of observations, y_i , that can be described by model functions, $M_i(\mathbf{x})$, where \mathbf{x} is the vector of model parameters, we seek to find \mathbf{x} for which the sum

$$S = \sum_{i=1}^{n} w_i [y_i - M_i(\mathbf{x})]^2$$
 (8.3.2.1)

is minimum. For restrained refinement, S is composed of several classes of observational equations, including, in addition to the ones for structure factors, equations for interatomic distances, planar groups and displacement factors.

Structure factors yield terms in the sum of the form

$$\Delta_{\rm SF} = \left[|F_{\rm obs}(\mathbf{h})| - |F_{\rm calc}(\mathbf{h})| \right]^2 / \sigma_{\mathbf{h}}^2. \tag{8.3.2.2}$$

The distances between bonded atoms and between next-nearest-neighbour atoms may be used to require bonded distances and angles to fall within acceptable ranges. This gives terms of the form

$$\Delta_d = (d_{\text{ideal}} - d_{\text{model}})^2 / \sigma_d^2,$$
 (8.3.2.3)

where σ_d is the standard deviation of an empirically determined distribution of values for distances of that type. Groups of atoms may be restrained to be near a common plane by terms of the form (Schomaker, Waser, Marsh & Bergman, 1959)

$$\Delta_p = (\mathbf{m}_l \cdot \mathbf{r} - d_l)^2 / \sigma_p^2, \tag{8.3.2.4}$$

where \mathbf{m}_l and d_l are parameters of the plane, σ_p is again an empirically determined standard deviation, and \cdot indicates the scalar product.

If a molecule undergoes thermal oscillation, the displacement parameters of individual atoms that are stereochemically related must be correlated. These parameters may be required to be consistent with the known stereochemistry by assuming a model that gives a distribution function for the interatomic distances in terms of the individual atom parameters and then restraining the variance of that distribution function to a suitably small value. The variation with time of the distances between covalently bonded atoms can be no greater than a few hundredths of an ångström. Therefore, the thermal displacements of bonded atoms should be very similar along the bond direction, but they may be more dissimilar perpendicular to the bond. If we make the assumption that the atom with a broader distribution in a given direction is 'riding' on the atom with the narrower distribution, the variance of the interatomic distance parallel to a vector v making an angle $\theta(\mathbf{v}, i)$ with the direction of bond i is (Konnert & Hendrickson, 1980)

$$V_{\mathbf{v}} = \Delta_{\mathbf{v}}^{2} \cos^{2} \theta + (\Delta_{\mathbf{v}}^{4}/2d_{0}^{2})(\sin^{4} \theta - 6\sin^{2} \theta \cos^{2} \theta) + \dots,$$
(8.3.2.5)

where d_0 is the normal distance for that type of bond, $\Delta_{\bf v}^2=(\overline{u}_a^2-\overline{u}_b^2)$, and \overline{u}_a^2 and \overline{u}_b^2 are the mean square displacements parallel to ${\bf v}$ of atom a and atom b, respectively. The restraint terms then have the form $V_{\bf v}^2/\sigma_{\bf v}^2$. For isotropic displacement factors, these terms take the particularly simple form $(B_a-B_b)^2/\sigma_B^2$, but with the disadvantage that, when isotropic displacement parameters are used, the displacements cannot be suitably restrained along the bonds and perpendicular to the bonds simultaneously.

Several additional types of restraint term have proved useful in restraining the coordinates for the mean positions of atoms in macromolecules. Among these are terms representing non-bonded contacts, torsion angles, handedness around chiral centres, and noncrystallographic symmetry (Hendrickson & Konnert, 1980; Jack & Levitt, 1978; Hendrickson, 1985). Contacts between nonbonded atoms are important for determining the conformations of folded chain molecules. They may be described by a potential function that is strongly repulsive when the interatomic distance is less than some minimum value, but only weakly attractive, so that it can be neglected in practice, when the distance is greater than that value. This leads to terms of the form

8.3. CONSTRAINTS AND RESTRAINTS IN REFINEMENT

Table 8.3.2.1. Coordinates of atoms (in Å) in standard groups appearing in polypeptides and proteins; restraint relations may be determined from these coordinates using methods described by Hendrickson (1985)

		пенанской	i (1905)	
Ī	Main chain, l	inks and termin	al groups	
	Main			
	N	1.20134	0.84658	0.00000
	$C\alpha$	0.00000	0.00000	0.00000
		-1.25029	0.88107	0.00000
	0 -	-2.18525	0.66029	-0.78409
	C terminal			
		1.20006	0.04700	0.00000
	N	1.20006	0.84799	0.00000
	$C\alpha$	0.00000	0.00000	0.00000
		-1.26095	0.86727	0.00000
	0 -	-2.32397	0.27288	-0.29188
	O_t -	-1.15186	2.04837	0.35987
	N amino term	sinal		
			0.04650	0.00000
	N	1.20134	0.84658	0.00000
	Cα	0.00000	0.00000	0.00000
		-1.25029	0.88107	0.00000
	0 -	-2.18525	0.66029	-0.78409
	N formyl tern	ninal		
ı	N	1.19423	0.82137	0.00000
ļ	Cα	0.00000	0.00000	0.00000
		-1.24896	0.88255	0.00000
		-2.10649	0.78632	-0.90439
	O_t		-0.77877	
	C_t	2.33913	0.39064	-0.53355
	C_t	2.33913	0.39004	-0.33333
	N acetyl term	inal		
	N dectyr term	1.19423	0.82137	0.00000
	$C\alpha$	0.00000	0.00000	0.00000
		-1.24896	0.88255	0.00000
		-2.10649	0.78632	-0.90439
	O_t		-0.77877	-0.93569
	C_t 1	2.33913	0.39064	-0.53355
	$C_t^{'}2$	3.44659	1.39160	-0.63532
	trans peptide	link		
	Cα	0.00000	0.00000	0.00000
	C	0.57800	1.41700	0.00000
	0	1.80400	1.60700	0.00001
		-0.33500	2.37000	0.00000
	Cα	0.00000	3.80100	0.00000
	cis peptide lin	nk		
ļ	$C\alpha$	0.00000	0.00000	0.00000
	C	1.30900	0.79200	0.00000
ļ	Ö	2.38500	0.17600	0.00000
	N	1.23500	2.11000	0.00000
	Cα	0.00000	2.90700	0.00000
Į			2.70700	0.0000
ļ	trans proline			
ļ	$C\alpha$	0.00000	0.00000	0.00000
	C	0.57800	1.41700	0.00000
	0	1.80400	1.60700	0.00001
ļ		-0.33500	2.37000	0.00000
ļ	$C\alpha$	0.00000	3.80100	0.00000
Į		-1.80000	2.19600	0.00000
I	cis proline lin		0.00000	0.00000
J	Cα	0.00000	0.00000	0.00000
ı	C	1.30900	0.79200	0.00000
ı	O	2.38500	0.17600	0.00000
J	N	1.23500	2.11000	0.00000
ļ	$C\alpha$	0.00000	2.90700	0.00000
ļ	$C\delta$	2.45500	2.93900	0.00000
- 1				

Table 8.3.2.1 (cont.)

14010 0.0.2.1 (00141)						
Side chair Ala A	Side chains for amino acids					
Cβ	0.02022	-0.92681	1.20938			
Arg R						
$C\beta$	-0.02207	-0.93780	1.20831			
$C\gamma$	-0.09067	-0.23808	2.55932			
$C\delta$	-0.79074	-1.07410	3.57563			
$N\varepsilon$	-0.76228	-0.46664	4.89930			
Cζ	-1.57539	-0.83569	5.89157			
$N\eta 1$	-2.60422	-1.65104	5.68019			
Νη2	-1.38328	-0.33329	7.11065			
Asn N	0.04600	1 00504	1 12104			
$C\beta$	0.04600	-1.02794	1.12104			
Сγ	-0.15292	-0.42844	2.50080			
Οδ1	-0.39364	0.78048	2.63809			
Νδ2	-0.06382	-1.27086	3.52863			
Asp D	0.04600	1.00704	1 10104			
Cβ	0.04600	-1.02794	1.12104			
Cγ	-0.15292	-0.42844	2.50080			
Οδ1	-0.39364	0.78048	2.63809			
Οδ2	-0.06930	-1.21904	3.46540			
Cys C	0.01017	0.05000	1 10077			
Cβ	0.01317	-0.95892	1.18266			
Sγ	-0.07941	-0.15367	2.80168			
Gln Q	0.01601	0.00604	1 16400			
Cβ	-0.01691	-0.98634	1.16423			
Cγ	-0.08291	-0.32584	2.52866			
Cδ	-0.20841	-1.31760	3.65937			
Οε1	-0.48899	-2.49684	3.46331			
Νε2	-0.00450	-0.81846	4.87646			
Glu E	0.06	0.05/55				
Сβ	-0.06551	-0.87677	1.25157			
Cγ	1.15947	-1.71468	1.59818			
Сδ	1.40807	-2.90920	0.72611			
Οε1	0.92644	-3.06007	-0.38343			
Οε2	2.16269	-3.74330	1.27140			
Gly G (no	nonhydrogen at	oms)				
His H						
$C\beta$	-0.06434	-0.96857	1.20324			
$C\gamma$	-0.52019	-0.29684	2.46369			
Νδ1	0.26457	0.53405	3.22184			
Cε1	-0.46699	1.05500	4.19371			
Ne2	-1.69370	0.59727	4.09040			
Cδ2	-1.75570	-0.25685	3.02097			
Ile I	0.05:05	0.0=:::				
Сβ	0.03196	-0.97649	1.23019			
Cγ1	-0.83268	-2.22363	0.92046			
Cγ2	-0.39832	-0.28853	2.54980			
Cδ1	-0.77555	-3.32741	2.01167			
Leu L	0.00005	0.04411	1 20241			
Cβ	0.09835	-0.94411	1.20341			
Cγ	-0.96072	-2.02814	1.32143			
Cδ1 Cδ2	-0.89548 -0.73340	-2.98661 -2.79002	0.13861 2.62540			
Lys K						
$C\beta$	-0.03606	-0.92129	1.21541			
C_{γ}	1.19773	-1.81387	1.35938			
Cδ	1.05466	-2.77178	2.53242			
$C\varepsilon$	2.34215	-3.51295	2.82637			
Νζ	2.16781	-4.42240	3.98733			

8. REFINEMENT OF STRUCTURAL PARAMETERS

Table 8.3.2.1 (cont.)

		` /	
Met M			
	0.02044	0.06506	1 17716
Сβ	0.02044	-0.96506	1.17716
$C\gamma$	-1.00916	-2.05384	1.00286
$S\delta$	-0.77961	-3.24454	2.37236
$C\varepsilon$	-2.08622	-4.42220	1.97795
Phe F			
Сβ	0.00662	-1.03603	1.11081
C_{γ}	0.03254	-0.49711	2.50951
Cδ1	-1.15813	-0.12084	3.13467
Cε1	-1.15720	0.38038	4.42732
Cζ	0.05385	0.51332	5.11032
$C\varepsilon 2$	1.26137	0.11613	4.50975
Cδ2	1.23668	-0.38351	3.20288
Pro P			
Сβ	0.12372	-0.78264	1.31393
C_{γ}	0.89489	0.13845	2.22063
Cδ	1.87411	0.86170	1.30572
	1.0/411	0.00170	1.30312
Ser S			
	-0.00255	0.00014	1 17/70
Сβ		-0.96014	1.17670
Ογ	-0.19791	-0.28358	2.40542
Thr T			
$C\beta$	-0.00660	-0.98712	1.23470
Ογ1	0.04119	-0.14519	2.43011
$C_{\gamma}^{\prime}2$	1.12889	-2.01366	1.21493
-,-			
Trp W			
Cβ	0.02501	-0.98461	1.16268
$C\gamma$	0.03297	-0.36560	2.51660
Cδ1	-1.03107	0.15011	3.20411
$N\varepsilon l$	-0.62445	0.62417	4.42903
$C\varepsilon 2$	0.72100	0.41985	4.55667
Cζ2	1.57452	0.72329	5.60758
$C\eta 2$	2.91029	0.38415	5.45120
Cη3	3.37037	-0.23008	4.28944
Cε3	2.51952	-0.53303	3.24549
Cδ2	1.17472	-0.33303 -0.20516	3.37412
002	1.1/7/2	0.20310	3.3/712
Tyr Y			
	0.00470	0.05229	1.20778
Cβ		-0.95328	
Сγ	-0.18427	-0.27254	2.54372
Cδ1	0.89731	0.26132	3.25049
$C\varepsilon l$	0.72371	0.85064	4.50059
Cζ	-0.54776	0.88971	5.06861
$\tilde{\text{C}}$ 2	-1.63905	0.38287	4.37622
Cδ2	-1.44975	-0.19374	3.12415
$O\eta$	-0.76405	1.40409	6.31652
5.,	0.70105	1.1010)	0.51052
Val V			
$C\beta$	0.05260	-0.99339	1.17429
	-0.13288	-0.39539 -0.31545	2.52668
Cγ1			
$C\gamma 2$	-0.94265	-2.12930	0.99811

$$\Delta_n = (d_{\min} - d_{\text{model}})^4 / \sigma_n^4,$$
 (8.3.2.6)

which are included only when $d_{\rm model} < d_{\rm min}$. Macromolecules usually gain flexibility by relatively unrestricted rotation about single bonds. There are, nevertheless, significant restrictions on these torsion angles, which may, therefore, be restrained by terms of the form

$$\Delta_t = (\chi_{\text{ideal}} - \chi_{\text{model}})^2 / \sigma_t^2, \tag{8.3.2.7}$$

Table 8.3.2.2. Ideal values for distances (Å), torsion angles (°), etc. for a glycine-alanine dipeptide with a trans peptide bond; distance type 1 is a bond, type 2 a next-nearest-neighbour distance involving a bond angle

Interatomi	c distance	s				
Number		S			Distance	Type
1	N(1)	to	C(1)α	,	1.470	1
2	$C\alpha(1)$	to	C(1)	,	1.530	1
3	C(1)	to	O(1)		1.240	1
4	N(1)	to	C(1)		2.452	2
5	$C(1)\alpha$	to	O(1)		2.432	2
6					1.469	1
7	N(2)	to	$C(2)\alpha$;	1.530	1
8	$C(2)\alpha$	to	C(2)		1.330	1
9	C(2)	to	O(2)			
	N(2)	to	C(2)		2.461	2
10	$C(2)\alpha$	to	O(2)		2.358	2
11	$C(2)\beta$	to	$C(2)\alpha$!	1.524	1
12	$C(2)\beta$	to	C(2)		2.515	2
13	$C(2)\beta$	to	N(2)		2.450	2
14	C(2)	to	$O(2)_t$		1.240	1
15	O(2)	to	$O(2)_t$		2.225	2
16	$C(2)\alpha$	to	$O(2)_t$		2.377	2
17	N(2)	to	C(1)		1.320	1
18	N(2)	to	O(1)		2.271	2 2
19	N(2)	to	$C(1)\alpha$!	2.394	2
20	$C(2)\alpha$	to	C(1)		2.453	2
Planar gro	une					
1	CTRM	$C(2)\alpha$	C(2)	O(2)	O(2)	
2	LINK	$C(2)\alpha$ $C(1)\alpha$	C(2) C(1)	O(2) $O(1)$	N(2)	$C(2)\alpha$
2	LINK	$C(1)\alpha$	C(1)	O(1)	11(2)	$C(2)\alpha$
Chiral cen	tres					
						Chiral
		Central				volume
		atom				(\mathring{A}^3)
1	Ala	$C(2)\alpha$	N(2)	C(2)	$C(2)\beta$	2.492
D 11	1 1 1					
Possible n		contacts			ъ.	
Number			0(1)		Distance	
1	N(1)	to	O(1)		3.050	
2	N(2)	to	O(2)		3.050	
3	O(2)	to	$C(2)\beta$		3.350	
4	N(2)	to	$O(2)_t$		3.050	
5	$O(2)_t$	to	$C(2)\beta$	3	3.350	
Torsion angles						
N(1)	$C(1)\alpha$	C(1)	N(2)		0.0	
$C(1)\alpha$	C(1)	N(2)	$C(2)\alpha$		180.0	
$C(1)\alpha$	N(2)	$C(2)\alpha$	C(2)a	,	0.0	
N(2)	$C(2)\alpha$	$C(2)\alpha$	$O(2)_t$		0.0	
14(2)	C(2)u	C(2)	$O(2)_t$		0.0	

where χ_{ideal} and χ_{model} are dihedral angles between planar groups at opposite ends of the bond.

Interatomic distances are independent of the handedness of an enantiomorphous group. If \mathbf{r}_c is the position vector of a central atom and \mathbf{r}_1 , \mathbf{r}_2 , and \mathbf{r}_3 are the positions of three atoms bonded to it, such that the four atoms are not coplanar, the *chiral volume* is defined by

$$V_c = (\mathbf{r}_1 - \mathbf{r}_c) \cdot [(\mathbf{r}_2 - \mathbf{r}_c) \times (\mathbf{r}_3 - \mathbf{r}_c)], \tag{8.3.2.8}$$

where \times indicates the vector product. The chiral volume may be either positive or negative, depending on the handedness of the group. It may be restrained by including terms of the form

$$\Delta_c = (V_{\text{ideal}} - V_{\text{model}})^2 / \sigma_c^2.$$
 (8.3.2.9)

Table 8.3.2.1 gives ideal coordinates, in an orthonormal coordinate system measured in Å, of various groups that are

8.3. CONSTRAINTS AND RESTRAINTS IN REFINEMENT

Table 8.3.2.3. Typical values of standard deviations for use in determining weights in restrained refinement of protein structures (after Hendrickson, 1985)

$\sigma_d = 0.02 \text{ Å} \\ 0.03 \text{ Å} \\ 0.05 \text{ Å} \\ 0.05 \text{ Å}$
$\sigma_p = 0.02 \text{ Å}$
$o_p = 0.02 R$
$\sigma_c = 0.15 \text{ Å}^3$
t
$\sigma_n = 0.50 \text{ Å}$
$\sigma_t = 15^{\circ}$ 3°
3° 15°
15°
Isotropic
$\sigma_B = 1.0 \text{ Å}^2$
1.5 Å^2
1.5 Å^2
2.0 Å^2

commonly found in proteins. The ideal conformations of pairs of amino acid residues, from which the ideal values to be used in restraint terms of various types may be determined, are constructed by combining the coordinates of the individual groups. For example, consider a dipeptide composed of

glycine and alanine joined by a trans peptide link, giving the molecule

$$\begin{array}{cccc} O(1) & C\beta(2) & O(2) \\ \parallel & \parallel & \parallel \\ N(1)-C\alpha(1)-C(1)-N(2)-C\alpha(2)-C(2)-O_t(2). \end{array}$$

The origin is placed at each of the $C\alpha$ positions in turn, and interatomic distances to nearest and next-nearest neighbours are computed. Planar groups and possible nonbonded contacts are identified, and torsion angles and chiral volumes for chiral centres are computed. Table 8.3.2.2 is a summary of the restraint information for this simple molecule. In order to incorporate this information in the refinement, these ideal values are combined with suitable weights. Table 8.3.2.3 gives values of the standard deviations of the various types of constraint relation that have been found (Hendrickson, 1985) to give good results in practice.

Even for a small protein, the normal-equations matrix may contain several million elements. When stereochemical restraint relations are used, however, the matrix elements are not equally important, and many may be neglected. Convergence and stability properties can be preserved when only those elements that are different from zero for the stereochemical restraint information are retained. The number of these elements increases linearly with the number of atoms, and is typically less than 1% of the total in the matrix, so that sparse-matrix methods (Section 8.1.5) can be used. The method of conjugate gradients (Hestenes & Stiefel, 1952; Konnert, 1976; Rae, 1978) is particularly suitable for the efficient use of restrained-parameter least squares.